

22

23 **Supplement B – Model Documentation**¹

24 **CSFMM – Classical Swine Fever Management Model**

25 The following documentation details the structure of the model as implemented in C++
26 programming language. The description orients to the ODD protocol (Overview, Design,
27 Details) for documentation of complex models (Grimm et al., 2006. A standard protocol for
28 describing individual-based and agent-based models. *Ecol.Modell.* 198, 115-126) and extends
29 the OD parts already provided in the manuscript. Description focuses on the model version as
30 applied to the presented simulations – a full featured description of the CSFMM may be
31 found at (EcoEpi, 2011).

32 ***Purpose***

33 The CSFMM was developed to evaluate different aspects of CSF-outbreak management by
34 simulating the geographical spread of the infection on a “landscape” of pig herds.
35 The formulation of model rules on how the disease spreads through an area as well as on how
36 different management measures interfere with this spread is determined by the existing expert
37 concepts. The simulation tool collates the relevant expert rules on the very grainy basis thus
38 the outbreak dynamics seen in the model are emerging bottom-up from the conceptual model
39 input. The main purpose of the modelling initiative is to test suggested management concepts
40 or control schemes given the state-of-the-art knowledge on the disease.

41 ***Overview***

42 The CSFMM is a spatially explicit, stochastic, state-transition model comparable to several
43 similar models (Bates et al., 2003; Garner and Beckett, 2005; Harvey et al., 2007).

44 The model construction orients to the approach of the more generic NAADSM (Harvey et al.,
45 2007). The description was prepared to parallel the model documentation of these authors to
46 enhance comparability although the CSFMM was not a reimplementation of the former and
47 its focus is exclusively on CSF management problems.

48 The model is organised in an object-oriented manner to enhance efficient alterations
49 according to simulation protocols on the level of control activities and their interaction.

50 In CSFMM, disease spread occurs between pig herds at specified locations, and is dependent
51 on relative locations and distances between herds. After introduction of the virus into the
52 herd, the infection follows a predictable cycle over time, moving the herd from one disease
53 state to the next. This cycle may be interrupted by intervention of disease control
54 mechanisms. Stochastic decisions drive all processes within the model for which empirical
55 probability distributions are reported (i.e. detection, spread, testing).

56 The model reflects an administrative border between two halves of the simulation area that
57 allow for alternative cooperation scenarios with regards to the adjustment of control and
58 mitigation measures in both subareas.

59 *The components and input parameters of the model are described in the following sections:*
60 *Section organisation, Section disease, Section spread, Section disease detection, Section*
61 *tracing out, Section control measures, Section priorities of actions and Section costs.*

¹ Version with different densities and different management practice on to sides of an assumed border
(Documentation derived from template: Version 8.0 October 2010, see EcoEpi 2011)

62 **Organisation**

63 **Entities, state variables, scales**

64 The CSFMM was formulated with two hierarchical entities: herd and landscape.

65

66 The low-level entities are individual herds and the transmission of the infection is followed on
67 the herd level (cf. units in (Harvey et al., 2007)). Animals inside a herd are reflected by
68 numbers and within herd spread follows deterministic epidemic dynamics (e.g. (Klinkenberg
69 et al., 2003)).

70 The state variables of the herd entities that are stochastically pre-assigned during initialisation
71 are its geographical position (x,y coordinates; random distribution), and its size (stocking
72 number; see eq.1). Additionally, each herd is assigned an initial age (time since last
73 restocking).

74 Disease state variables reflect the herd's CSF status (susceptible, infected, infectious), and
75 whether it is involved in the outbreak (detected or not detected, number of infected or sero-
76 converted animals) or any control activity (standstill zone, surveillance zone, pre-emptive
77 culling, vaccination, tracing or testing).

78 The herd size is assumed to be static: herd size is not altered by the movement of animals.
79 Only when a herd is destroyed or not restocked the number of animals will be affected (i.e.
80 herd gets empty). Finished herds [**Par**: DaysNeededTillSell] are instantaneously restocked
81 assuming slaughter of all pigs and disinfection i.e. the age and infection features are reset.

82

83 The second level entity is the landscape (simulation area) that maintains information on the
84 whole population, the density and the actual stage of control measures (i.e. outbreak detected,
85 or final success). The landscape comprises two subareas arranged horizontally above and
86 below a line that represents an administrative border. Both subareas of the landscape are
87 independently initialised at the beginning of each simulation by distributing a number of
88 herds per sq km [**Par**: HerdDensity] and the realisation of an average herd size [**Par**:
89 PigsPerHerd] following the distribution:

90

91 eq.1 Herd size = PigsPerHerd/4 + random number between 0 and 1.5 * PigsPerHerd
92 (e.g.: if PigsPerHerd=1000; then herd size is between 250 and 1750)

93

94

95 **Scheduling**

96 The model steps forward in time by one day. On each simulation day, a regular sequence of
97 processes is scheduled by the landscape:

98

Init: Generate farm landscape (herds with age, position, size)

Update Status: Increment of the disease and vaccination status of each single herd.

Pass Infection: Evaluate potential transmission to next herds (regional and local)

Detect Infection: Test every infected herd whether detected today

Sell: Handle finished finisher herds (welfare or pre-tested slaughter);

Make Priorities: Organise scheduling of control measurements (culling & testing);

Start Measure: Perform control measures along the schedule up to daily capacity;

Check Stop Condition: Eradication; or maximum number of control days

Stop: Output and Statistics

99
100
101
102
103
104

Process operations are performed synchronously for all related herds (i.e. resulting changes in status of a herd does not affect the process itself during one day).
At the end of the simulation run the overall statistics are prepared.

105 Design Concepts

106 Emergence: The model is designed and parameterised to allow emergence of dynamics on the
107 level of epizooty from simple and expert approved epidemiological details – rule-based
108 modelling approach.

109
110 Stochasticity: Most dynamic processes are modelled stochastic. The reason is high uncertainty
111 about very particular realisations for e.g. herd assemblages, daily disease transmission
112 pathway and temporal order, detection distribution, traceability of the infection network,
113 human compliance with regulations.

114
115
116

Table 1: Overview of stochastic components of the CSFMM

Process/Mechanism	Distribution	Parameter(s)
Locations of herds	2D uniform(0;p)	p: AreaSize (per subarea)
Stocking size (eq.1)	Uniform(1/4*p; 7/4*p)	p: PigsPerHerd (per subarea)
Detection by clinics	Uniform(p _u ;p _o)	p _u : MinFarmerDetectTime p _o : MaxFarmerDetectTime
Regional transmission		
- Distance (eq.2b)	Negative Exponential(p ₁) Censored at p ₂	p ₁ : RegionalMeanInfectDist p ₂ : RegionalMaxInfectDist
- Infection given contact	Bernoulli(p)	p: RegionalInfectProb
Movement ban		

- Standstill effect	Bernoulli(p)	p: StandstillEfficiency
Local infection		
- up to 500m	Bernoulli(p ₁)	p ₁ : D500
- up to 1000m	Bernoulli(p ₂)	p ₂ : D1000
Traceability of infection network	Bernoulli(p)	p: TracingEfficiency

117

118 **Submodels (ordered by schedule)**

119 ***Disease***

120 **Disease dynamics within the basic entity (herd)**

121 When a susceptible herd is infected, it transits instantaneously to incubating, and further to
 122 infectious according to the parameter describing herd incubation [**Par**: HerdIncubDays]; e.g.
 123 when an infected herd is assumed to be able to cause infections itself. When infected herd
 124 switches to infectious the infectious animal population of the herd is seeded randomly with 2-
 125 4 infectious pigs. Further growth of the infectious respective recovered cohort follows
 126 standard exponential growth with random increments [**Par**: EpidemicGrowthRate], i.e. for
 127 each virus positive pig one comes up with daily rate of 0.0822 (parameterised according to
 128 (Klinkenberg et al., 2003)).

129 The course of the disease in an infected herd is not altered by re-infection (i.e. no super-
 130 infections are modelled). Technically, a herd that receives an infection (e.g. by animal
 131 transport) could be regarded as immediately infectious. Treating newly infected herds as
 132 incubating, however, reflects the fact that most of the animals in the herd still need to progress
 133 through the early state of the infection.

134 **Transmission**

135 According to the literature the model differentiates two modes of CSF transmission (Staubach
 136 et al., 1997;Stegeman et al., 2002;Ribbens et al., 2004): Distance dependent or local
 137 transmission refers to all processes that cause decreasing risk by distance in the vicinity of an
 138 outbreak herd (e.g. insects, children, farmer's movement etc.) and is usually attributed to
 139 surroundings up to 500-1000m (Staubach et al., 1997;Stegeman et al., 2002). Distance
 140 independent or regional transmission refers to routes by which the risk of transmission does
 141 not depend on this distance (Lorry or service personal moving around (Ribbens et al., 2004)).
 142 Any infected herd that is no longer incubating is assumed infectious i.e. is capable of
 143 spreading disease.

144 Local transmission:

145 Local transmission summarizes distance dependent transmission events. These are associated
 146 to any neighbourhood transportation of the virus making the infection risk continuously
 147 decreasing by distance from the source.

148 According to the analysis by Stegeman et al. (2002) two parameters for local transmission are
 149 used (Table 1): An infectious herd passes the infection either of its neighbours according to a
 150 Bernoulli probability up to 500m [**Par**: D500] and further up to 1000 meters [**Par**: D1000].
 151 Local transmissions are recorded as such but cannot be identified during tracing
 152 investigations.

153 Regional transmission:

154 Regional transmission simulates distance independent virus transport (Staubach et al., 1997)
155 or “direct contact” spread (Harvey et al., 2007). The scale of the transmission events is
156 “regional” i.e. they may cover large parts of the pig area without decreasing final risk of
157 infection.

158 The regional transmission occurs from an infectious source herd according to a twofold
159 random procedure: First, on each simulation day, a distance is randomly drawn from the
160 regional distance distribution, i.e. negative exponential with specified mean [**Par**:
161 RegionalMeanInfectDist], and up to a specified maximum distance [**Par**:
162 RegionalMaxInfectDist] but beyond 1000m. Distances are calculated according to the
163 Euclidean metric.

164

165 eq.2b Distance = negexp(P->RegionalMeanInfectDist); on $1 \leq \text{Dist.} \leq \text{RegionalMaxInfectDist}$

166

167 With the resulting distance a geographic position of a recipient herd is searched that is closest
168 by the randomly drawn distance. If necessary, the recipient is randomly selected out of
169 multiple, evenly matching candidates.

170 If the recipient herd is not susceptible, or was already destroyed, the contact does not occur. If
171 the recipient herd is susceptible the model randomly determines whether an infectious contact
172 happens based on the regional transmission probability [**Par**: RegionalInfectProb]. Then the
173 recipient status becomes infected.

174 Successful transmission events are recorded and can be identified later during tracing
175 investigations. The number of animals in a movement is not considered (Harvey et al., 2007).

176 **Detection**

177 Infected herds are detected by routine check for clinical suspicion, routine testing according to
178 surveillance schemes, or targeted testing after successful establishment of a contact by tracing
179 investigations or diagnostic protocols.

180 There are no false-positive detections in model.

181 The order of potential detection in the model is:

182 D1) Tracing related tests (diagnostic test; census or sample)

183 D2) Visit according to surveillance scheme (vet)

184 D3) Otherwise routine daily check (farmer)

185 Only infectious herds are detected by D2 and D3. D1 can detect even earlier according to the
186 rtRTPCR test characteristics. After detection the herd is designated as detected and measures
187 that are dependent on detection are initiated subsequently, e.g. tracing of backward and
188 forward contacts will be schedule for the next day. Dependent on border scenario first
189 detection is handled independent for both subareas of the landscape.

190 **Basic detection**

191 If a herd gets infected a regular day post infection is fixed for detection. The value is
192 randomly drawn of the interval [**Par**: MinFarmerDetectTime; **Par**: MaxFarmerDetectTime].

193 DayOfFarmerDetection = today + MinFarmerDetectTime

194 + integer[uniform[0,1] * (MaxFarmerDetectTime - MinFarmerDetectTime)];

195 The standard interval is parameterized according to Dutch outbreak data (Klinkenberg et al.,
196 2005) and covers MinFarmerDetectTime = 21 and MaxFarmerDetectTime = 55 days post
197 infection. The resulting distribution reread from the model generally mimics the data while
198 particularities might change with outbreak realisation.

199 Any process that is supposed to alter detection modulates the regular detection date per herd.

200 For example, as long as the first notification is missing the respective detection date per herd

201 is shifted later by 14 days [**Par**: FirstDetectDelay] (Stegeman et al., 1999b). Thus, during the
202 high risk period the detection happens between day 35 and 69 (i.e. 5th to 10th week post
203 infection; see (Fritzemeier et al., 2000))

204 **Farmer**

205 Every day the model tests if a herd has reached the detection date. These notifications relate to
206 the successful confirmation of the disease after farmers suspicion.

207 **Veterinary Service**

208 After outbreak notification inside of designated surveillance zones once a week a targeted
209 visit by an expert is assumed that shortens time till detection by a number of days [**Par**:
210 SurveillanceDetectReduction].

211 **Tracing**

212 If an infected herd was detected, next day tracing happens along the recorded transmission
213 history. Tracing might establish the source contact (one step back; tracing backward) or any
214 herd infected by the detected one (multiple one step forward; tracing forward). The actual
215 identification of these links during tracing investigations is reflected by a probability value i.e.
216 the tracing efficiency [**Par**: Forward/BackwardTracingEfficiency]. For example, assuming
217 100% tracing efficiency will guarantee the stepwise but full reconstruction of all regional
218 transmission events. Traced herds are marked for diagnostic testing. Herds could be involved
219 in multiple tracing investigations. Tracing only works if animals are present. If the herd traced
220 next is already culled the tracing is terminated on that branch. Dependent on border scenario
221 tracing of cross-border infection events may be possible or excluded.

222 **Diagnostic Testing**

223 Diagnostic testing is simulated whenever the strategy requires diagnostic tests as basis for
224 decisions on a herd status. Possible examples are: infected herds designated for testing from a
225 traced link, herds designated for pre-emptive destruction, or final screening before the lift-up
226 of restrictions. The diagnostic failures in confirmatory tests, e.g. after farmer suspicion, are
227 not modelled explicitly but covered by the basic detection period. The model test system
228 comprises of the following features:

229 Test target: virus positive test units [**Par**: TestSystem],

230 Test sensitivity: the usual probability to detect a truly positive test unit [**Par**: TestSensitivity]

231 Test operability: the time period necessary after infection until the test will notice CSF
232 infections [**Par**: TestDaysTillDetect]

233 Test sample: The sampling in a tested herd covers all animals (census) [**Par**: TestSample]

234 **Selling/Restocking**

235 For all herds that are not yet detected as infected and are not emptied, the time since simulated
236 last restocking is checked against the maximum fattening age [**Par**: DaysNeededTillSell]. If
237 maximum fattening age was reached the slaughter and immediate restocking is simulated by
238 resetting of disease state and age. If the CSF outbreak in the simulation area was already
239 detected, slaughter was completely excluded due to movement restrictions and the aging herd
240 finally will be destroyed for welfare reasons.

241 **Intervention measures**

242 Intervention measures refer to the establishment of surveillance and standstill zones. Before
243 first notification none of these measures is activated in the model. Dependent on border
244 scenario simulated zones overlap or are cut along the borderline.

245 **Surveillance zone**

246 The circular area [**Par**: SurveillanceRadius] established around infected herds yet detected.
247 Following detection of the centre herd the zone remains activated at least as long as strategy
248 concept foresees [**Par**: SurveillanceDuration]. Subsequent detections during simulation might
249 cause prolonged effective duration.

250 The intervention measure causes increased awareness e.g. by regular veterinary inspection.
251 Thus likely more timely detection of an infected herd is based on an additional stochastic test
252 once per week against a reduced basic detection time [**Par**: SurveillanceDetectReduction] (see
253 chapter Detection).

254 **Standstill zone**

255 The circular area [**Par**: StandstillRadius] established around infected herds yet detected. The
256 zone is activated after time necessary for legislative process [**Par**: StandstillDelay]. Following
257 detection of the centre herd the zone remains activated at least as long as strategy concept
258 foresees [**Par**: StandstillDuration]. Subsequent detections during simulation might cause
259 prolonged effective duration. This happens if any herd already in standstill zone A, became
260 contained in a further newly established standstill zone B. In consequence, the standstill time
261 [**Par**: StandstillDuration] is restarted also for the complete previous standstill zone A.

262 The intervention measure aims at halting of all animal movements. Thus, regional (or
263 distances independent) transmission events initiated by the transmission process are
264 suppressed with certain efficiency [**Par**: StandstillEfficiency] for contacts originating from or
265 connect into active standstill zones. Any herd that becomes emptied by control measures will
266 not be restocked if it is assigned to a standstill zone. Standstill is assumed not to influence the
267 local neighbourhood of the centre herd which is intended to result in less favourable outcome
268 for any strategy that does not apply control measures to the core neighbourhood of an
269 outbreak herd. The basic motivation was inconsistent knowledge. Certain contact in the close
270 vicinity of an outbreak, however, should also be excluded by movement ban in practice.

271 After lift-up of standstill restriction in an area all empty herds are assumed to be restocked
272 immediately (Mangen et al., 2002).

273 **Control measures**

274 Control measures simulated by CSFMM are stamping-out and pre-emptive culling. Before
275 first notification none of these measures is activated in the model.

276 **Destruction**

277 Immediately with the infected herd that is detected first, the destruction program starts. There
278 are three cases where herds are destroyed in the model: stamping out of infected herds after
279 detection, pre-emptive culling in a circular neighbourhood of detections (i.e. ring destruction),
280 and welfare slaughter inside of standstill zones.

281 There is a limit to the number of animals that can be destroyed per day. This is referred to as
282 the culling capacity [**Par**: CullCapacityPerDay]. Herds designated for destruction queue by
283 priority (see list below). Every day queuing herds are culled upon saturation of the daily
284 capacity. Queuing herds are not quarantined.

285 Stamping-out:

286 In the model all detections of infected herds are destroyed after a number of days reserved for
287 preparation [**Par**: CullDelay]. Herds designated for stamping-out are assigned with the
288 highest priority for destruction. Hence, they instantaneously will be destroyed. Therefore, if
289 on one day the capacity limit is already reached by necessary stamping-out, it is assumed that
290 ad hoc destruction capacity will become available to allow instantaneous destruction of all
291 detected infected herds.

292 Pre-emptive culling:

293 Subsequent to stamping-out all herds in a circular area around the detected herd may be
294 designated for pre-emptive culling in accordance with the simulated strategic concept. The
295 size of the control area is defined by a radius [**Par**: CullRadius]. The culling zone is cleared
296 by concentric distances from the centre either from inner to outer herds or vice versa. Herds
297 designated for pre-emptive culling measures have second highest priority in the waiting list.

298 Welfare slaughter:

299 Finisher herds in standstill zone that reach fattening age remain in the model for an additional
300 of number of days [**Par**: SelldayExtension]. If then the standstill has still not been lifted these
301 herds are destroyed and not restocked. Welfare slaughter designation results in the lowest
302 priority for culling.

303 **Initialisation**

304 On the squared simulation area [**Par**: AreaSize] two horizontal subareas are determined (i.e.
305 AreaSize * 0.5 AreaSize). Independently for both subareas herds are distributed randomly
306 according to herd density parameterization [**Par**: HerdDensity]. Herds are populated
307 according to mean herd size parameter [**Par**: PigsDensity] randomly drawn between 26%-
308 175% the parameter value. Each herd is randomly assigned with a fattening age up to a
309 maximum [**Par**: DaysNeededTillSell]. The herd in the lower subarea that is closest to the
310 geometric point horizontally centred and 10km apart of the borderline is seeded as infected.

311 **References**

- 312 1. Bates, T.W., Thurmond, M.C., Carpenter, T.E., 2003. Description of an epidemic
313 simulation model for use in evaluating strategies to control an outbreak of foot-and-
314 mouth disease. Am. J. Vet. Res. 64, 195-204.
- 315 2. EcoEpi. CSFMM: Classical Swine Fever Management Model, Online Repository.
316 <http://www.ecoepi.eu/CSFMM> . 2011. 26-6-2011.
- 317 3. Fritzemeier, J., Teuffert, J., Greiser-Wilke, I., Staubach, C., Schlüter, H., Moennig, V.,
318 2000. Epidemiology of classical swine fever in Germany in the 1990s. Vet Microbiol
319 77, 29-41.
- 320 4. Garner, M.G., Beckett, S.D., 2005. Modelling the spread of foot-and-mouth disease in
321 Australia. Aust Vet J 83, 758-766.
- 322 5. Harvey, N., Reeves, A., Schoenbaum, M.A., Zagmutt-Vergara, F.J., Dubé, C., Hill,
323 A.E., Corso, B.A., McNab, W.B., Cartwright, C.I., Salman, M.D., 2007. The North
324 American Animal Disease Spread Model: A simulation model to assist decision making
325 in evaluating animal disease incursions. Prev. Vet. Med. 82, 176-197.

- 326 6. Klinkenberg, D., Everts-van der Wind, A., Graat, E.A.M., de Jong, M.C.M., 2003.
327 Quantification of the effect of control strategies on classical swine fever epidemics.
328 *Math. Biosci.* 186, 145-173.
- 329 7. Klinkenberg, D., Nielen, M., Mourits, M.C.M., de Jong, M.C.M., 2005. The
330 effectiveness of classical swine fever surveillance programmes in The Netherlands.
331 *Prev. Vet. Med.* 67, 19-37.
- 332 8. Ribbens, S., Dewulf, J., Koenen, F., Laevens, H., de Kruif, A., 2004. Transmission of
333 classical swine fever. A review. *Veterinary Quarterly* 26, 146-155.
- 334 9. Staubach, C., Teuffert, J., Thulke, H.-H., 1997. Risk analysis and local spread
335 mechanisms of classical swine fever. *Epidémiol. santé anim.* 31-32, 6.12.1-3.
- 336 10. Stegeman, A.J., Elbers, A.R.W., Bouma, A., de Jong, M.C.M., 2002. Rate of inter-herd
337 transmission of classical swine fever virus by different types of contact during the 1997-
338 8 epidemic in The Netherlands. *Epidemiol. Infect.* 128, 285-291.
339